

N-acetyl-cysteine in the treatment of Parkinson's disease. What are we waiting for?

Marcos Arturo Martínez-Banaclocha*

Servicio de Patología y Oncología Diagnóstica, Hospital Lluís Alcanyis, Xàtiva, Spain

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ABSTRACT

Parkinson's disease is an age-related neurodegenerative disorder that is ameliorated with levodopa. However, long-term use of this drug is limited by motor complications, postural instability and dementia resulting in the progression of the disease. Insights into the organization of the basal ganglia and knowledge of the mechanisms responsible for cell death in Parkinson's disease has permitted the development of putative neuro-protective drugs that might slow the disease progression. Although no drug has yet been established to alter the rate of disease progression, recent publications have confirmed previous results and hypotheses about the probable role of thiolic antioxidants on Parkinson's disease, demonstrating a significant reduction of dopaminergic neuronal degeneration in α -synuclein over expressing mice treated with oral N-acetyl-cysteine. This thiolic antioxidant is a modified form of the natural amino acid cysteine, which is the precursor of the most potent intracellular antioxidant glutathione. Besides, increasing evidence has been accumulated in the last 10 years about the beneficial effects of this thiolic antioxidant in experimental and pathologic states of the nervous system, including against neurotoxic substances. The present paper put forward the existing rationale evidence for the use of N-acetyl-cysteine alone or in combination with levodopa in the clinical management of this neurodegenerative disorder.

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Introduction

Parkinson's disease (PD) is a very common neurodegenerative disorder caused by idiopathic degeneration of dopamine-producing cells in the pars compacta of the substantia nigra located in the midbrain [1]. This specific neuro-degeneration leads to clinical signs including tremor at rest, rigidity on passive movement, bradykinesia, and hypokinesia. Postural instability, orthostatic hypotension, and dementia are invariably developed with the progression of the disease. Understanding the pathogenesis of the disease has been advanced in the last decade, being necessary to improve and found new therapeutic strategies based on scientist evidence.

Levodopa (L-dopa) is the main pharmacological treatment for PD, but its use is limited by the development of motor fluctuations and drug-induced dyskinesias. Dopamine agonists are also used, either alone or in combination with L-dopa, acting directly on dopamine receptors and mimicking endogenous dopamine functions. Monoamine oxidase B (MAO-B) inhibitors act increasing dopamine levels in the basal ganglia by inhibiting dopamine catabolism. Catechol O-methyl transferase (COMT) inhibitors also inhibit the catabolism of dopamine, thereby extending the half-life of L-dopa. However, all PD patients ultimately require L-dopa for control their symptoms.

There are numerous unanswered questions regarding the therapeutic management of PD. The present paper put forward the existing rationale evidence for the use of N-acetyl-cysteine (NAC) alone or in combination with L-dopa in the medical management of this disorder (Fig. 1).

Etiologic factors in Parkinson's disease

The pathogenesis of PD seems to be multi-factorial including environmental factors that act on genetically susceptible individuals as they age [2,3]. A broad spectrum of both genetic and environmental factors have been suggested as contributing to the initiation and progression of PD, but aging is the single most important risk factor for this disorder and undoubtedly contributes to PD progression through its accumulative oxidative damage, decrease in antioxidant ability and impairment of mitochondrial bioenergetic capacity in the brain [4–10].

Many studies have examined the impact of environmental agents on the risk of PD. In fact, PD patients show abnormalities of oxidative phosphorylation that impair their mitochondrial energy metabolism increasing reactive oxygen species (ROS) generation, which closely resembles that attributable to 1-methyl, 4-phenyl, 1,2,3,6-tetrahydropyridine (MPTP) [11–13] but this impairment is apparently constitutive in origin [14,15]. Schapira and co-workers were the first to report that mitochondrial Complex I activity was selectively reduced in the substantia nigra of patients with PD [16]. The Complex I impairment is worse in more

* Address: Urbanización Tiro de Pichón 77, 46740 Carcagente, Valencia, Spain.

E-mail address: martinez_marben@gva.es

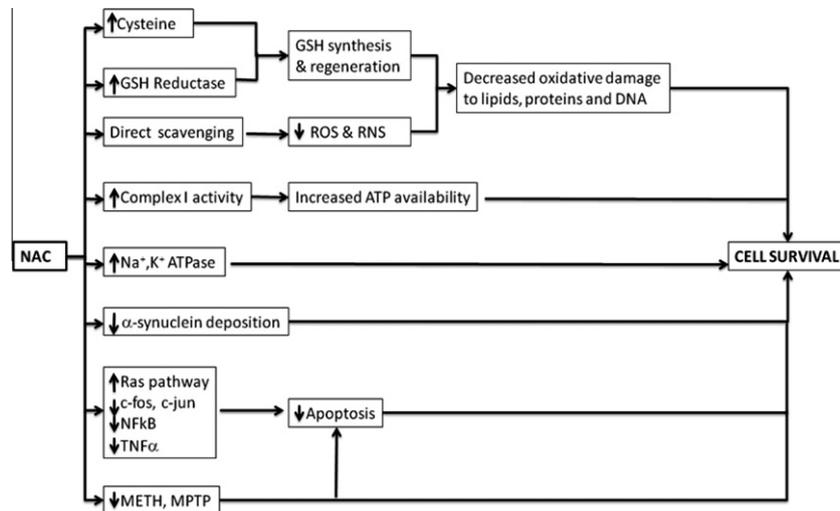


Fig. 1. Potential therapeutic actions of *N*-acetyl-cysteine (NAC) against dopaminergic neuro-degeneration.

advanced cases and seems to affect also non-nigral brain areas, muscle and fibroblasts of PD subjects [15,16].

Swerdlow's group carried out an elegant experiment using cybrids cells to confirm that the mitochondria in PD were at least 20% less efficient in Complex I activity, produced higher levels of ROS, and rendered their host cells more susceptible to MPTP-induced cell death [15]. They suggested that Complex I defect was in the mitochondrial DNA of PD patients caused by parental inheritance or by oxidative damage on the mitochondrial DNA. This constitutive defect in Complex I help to explain why some individuals develop PD following toxin exposure, while others do not. MPTP is not toxic by itself but into the brain it is transformed to the toxic product MPP+(1-methyl-4-phenylpyridinium) by MAO-B. MPP+ is selectively taken up by the dopaminergic neurons of the substantia nigra and selectively taken up by their mitochondria where it inhibits Complex I activity [13]. Then, a vicious cycle develops, which causes oxidative damage and bio-energetic deficiency into the neuron.

Six genes identified as α -synuclein (SNCA), ubiquitin C-terminal hydrolase like 1 (UCH-L1), parkin (PRKN), LRRK 2, PINK 1 and DJ-1 have been reliably linked to PD and/or to neurodegeneration of the parkinsonian type. These single gene mutations with the notable exception of LRRK 2 are responsible for only a small number of patients with PD. The LRRK 2 gene (PARK8) is the most common cause (5–7%) of familial PD to date [17].

Oxidative stress and glutathione deficiency

The earliest reported biochemical change identified in the substantia nigra of early PD patients is a significant depletion of reduced glutathione (GSH), which may promote morphological mitochondrial damage by ROS [18,19]. GSH (γ -L-glutamyl-L-cysteinyl-glycine) is the most abundant intracellular non-protein and water-soluble thiol antioxidant and it is synthesized by two-step reaction [20]. The mitochondrial GSH is dependent on the uptake from the cytosol since they lack the enzymes for the GSH synthesis [20]. GSH plays an important role in scavenging ROS and reactive nitrogen species (RNS) and in recycling other antioxidants and is kept in its thiol-reduced form (>98%) by glutathione disulfide (GSSG) reductase that maintains optimal GSH/GSSG ratios into the cell [20].

The magnitude of GSH depletion seems to parallel the severity of the disease and is the earliest known indicator of the substantia nigra degeneration preceding detectable losses in both mitochondrial

Complex I activity and striatal dopamine content [21]. Since Complex I impairment results in the generation of ROS [22] in agreement with reports of elevated markers of oxidative damage to lipids, proteins and DNA in the substantia nigra of patients with PD [23] and it has been demonstrated that exposure of mitochondrial membranes to nitric oxide resulted in selective and persistent inhibition of Complex I activity via S-nitrosation of critical thiol groups in the enzymatic complex, the inhibition of Complex I activity may be reversible by restoring mitochondrial GSH levels [24–26]. Besides, it has been shown that Complex I inhibition following prolonged dopaminergic GSH depletion in vitro was reversible with dithiothreitol, suggesting that it involved a reversible cysteine thiol modification. Then, it seems that GSH depletion in the substantia nigra of PD patients result in increased ROS and RNS generation leading to Complex I inhibition with subsequent mitochondrial dysfunction that significantly affects glutathione synthesis closing the vicious circle that ultimately leads to dopaminergic cell death [26].

In addition, oxidative stress may increase the accumulation of toxic forms of α -synuclein through oxidative ligation to dopamine playing a central role in PD [27,28], suggesting that increased oxidative stress due to early GSH deficiency in the substantia nigra may lead to enhanced toxicity of α -synuclein in dopaminergic neurons in PD.

NAC and Parkinson's disease

Having in account the accumulative oxidative damage in PD patients, some clinical studies have been performed using antioxidants in the treatment of PD with controversial results [29–31].

The central implication of GSH deficiency in PD has stimulated many investigations to find new potential approaches for maintain or restore GSH levels in these patients. Moreover, the use of GSH as a therapeutic agent is limited by its very short half-life in human plasma (<3 min) and difficulty to cross cell membranes, being necessary high doses to reach therapeutic levels [20,32]. Under physiological conditions, the cellular availability of cysteine is considered to be the rate-limiting factor in the synthesis of GSH. However, cysteine is toxic at high concentrations as the result of free radicals generation during cysteine autooxidation [33]. As a consequence, compounds that can be metabolized to cysteine could be used as pro-drugs to increase neuronal GSH levels.

NAC is the simplest cysteine pro-drug that can be systemically administered to deliver cysteine to the brain [8,34–37], acting as a

precursor for glutathione synthesis as well as a stimulator of the cytosolic enzymes involved in glutathione regeneration. Increase in Complex I activities *in vivo* and *in vitro* in mitochondria isolated from pre-synaptic terminals of aged mice was proposed 10 years ago as evidence that NAC was able to cross the blood brain barrier having reparative effects on brain mitochondria and against age-associated memory decline [9,34,35]. Since GSH levels become more depleted in the substantia nigra as the disease progresses, NAC may contribute to GSH repletion, which in addition to its potent antioxidant effects by direct scavenging of ROS can make this antioxidant ideal for counteract mitochondrial impairment in the substantia nigra of PD patients [38]. Furthermore, we have shown that NAC can prevent dopamine induced programmed cell death in cultured human cortical neurons [39] and also it can increase mitochondrial complex IV specific activity both *in vitro* and *in vivo* in synaptic mitochondrial preparations from aged mice [9,34].

Systemic administration of NAC increases brain levels of glutathione in mice [34,37,40,41], reduces markers of oxidative damage [38], increases brain synaptic [34,36] and non-synaptic brain [42] mitochondrial Complex I activities and protects against MPTP toxicity [43,44] and dopamine-induced cell death [39,45]. Besides, dietary supplementation of NAC during 1 year was able to counteract age-related decrease in rat brain expression of subunit 39 kDa and ND-1 of the mitochondrial respiratory Complex I and other subunits of the mitochondrial oxidative phosphorylation [46].

In view of the above, there are sufficient scientific evidence that Complex I inhibition by prolonged GSH depletion may be due, at least partially, to a reversible age-related event involving cysteine residues with impact on its enzymatic activity. This may be reversible by restoring GSH to normal levels and suggests that therapeutics toward the maintenance of cellular GSH concentration within dopaminergic neurons would be beneficial in PD. Moreover, *in vitro* studies have shown that NAC is able to restore Complex I age-related activity decline suggesting a direct role of NAC on cysteine residues [34,42].

On the other hand, recent studies using positron emission tomography suggested that chronic use of methamphetamine (METH) causes the reduction of dopamine transporter in the human brain, suggesting that this is the mechanism of neurotoxicity in humans [47]. These findings are supported by a report that demonstrates that the densities of dopamine transporter are significantly decreased in the postmortem striatum of chronic METH users [48]. Although the precise mechanisms of METH-induced neurotoxicity in dopaminergic nerve terminals are not fully known a recent positron emission tomography study demonstrated that NAC administration significantly attenuated the reduction of dopamine transporter in the monkey striatum 3 weeks after the administration of METH [49], possibly rescuing GSH levels in the striatum [50]. Therefore, it is likely that NAC would be a suitable substance for the treatment of neurotoxicity in dopaminergic nerve terminals related to the chronic use of METH in humans.

Oral NAC administration protected against loss of dopaminergic terminals associated with over-expression of α -synuclein in a mouse model [51]. The results of this study showed that striatal tyrosine hydroxylase positive terminal density was increased in NAC-treated α -synuclein over-expressing mice compared to α -synuclein over-expressing mice with a control diet. This also correlated with a decrease in α -synuclein immuno-labeling in the brains of over-expressing mice treated with NAC. Moreover, NAC supplementation significantly increased GSH concentrations in the substantia nigra of transgenic mice over-expressing α -synuclein [51].

There is growing evidence that NAC may play a role against programmed cell death (PCD) in postmitotic cells and oligodendrocytes *in vitro* [39,45,52,53]. Moreover, since alterations in mitochondrial structure and function are early events in apoptosis

[54] and NAC can prevent ROS accumulation, telomere shortening and cell death in an *in vitro* model that disrupt mitochondrial electron transport function it is conceivable that this thiolic antioxidant could act *in vivo* against PCD in PD [55]. NAC also inhibited the expression of *c-fos* and *c-jun* genes and TGF β -1 mediated apoptosis in human ovarian carcinoma cells [56]. In addition, long-term treatment with NAC affected NF-kappaB signaling in the brain of mice by increasing cytoplasmic retention of NF-kappaB thus preventing its action as a transcription factor in the nucleus [51]. Since increased activation of NF-kappaB may contribute to the pathology in models of Parkinson's disease, it is possible that NAC actions against modification of sulfhydryl groups in the proteins involved in regulating cell survival and NF-kappaB pathway were linked to reduced NF-kappaB activity in these models [57,58], being another beneficial action of NAC.

NAC can also inhibit TNF α -induced PCD in human neuronal and U937 cells by the preservation of mitochondrial integrity and function since NAC was able to partially prevent the mitochondrial membrane depolarization induced by this cytokine [39,59,60].

One study showed that NAC is a potent scavenger of both H₂O₂ and toxic quinones derived from dopamine and it prevented also dopamine-mediated inhibition of Na⁺, K⁺-ATPase activity suggesting another mechanism for the use of NAC in the treatment of PD [61]. Then, NAC may act against Na⁺, K⁺-ATPase inhibition, counteracting intracellular damage pathways that lead to death of dopaminergic neurons.

Recommendations for future research

They are necessary double blind randomized, placebo and well-designed and controlled clinical studies for test the probable benefit of oral NAC administration in ameliorate the symptoms and slow down the progression of PD. The age of onset of the symptoms can be important in order to classify the patients in two or even three sub-groups that will receive NAC. Standardization of patients' variables, doses of NAC used and reporting results will facilitate inter-study comparisons.

The reliability and validity of the Unified Parkinson Disease Rating Scale (UPDRS) has been widely documented, and it is currently the most common instrument used to measure the progression of PD. Investigators should report baseline, endpoint, and change in UPDRS scores, along with their respective standard deviations, since some researchers only report the motor sub-score in their investigations. It will be important that the activities of daily living (ADL) score be reported as well. Investigators should make sure to power their efficacy studies appropriately in order to make conclusive findings.

Given that most patients may be under actual treatments with L-dopa into, and given that an important treatment outcome is whether an additional drug allows for a decrease in L-dopa dose, data regarding actual L-dopa doses are quite important in the evaluation of NAC benefit.

While PD is mainly a disease of the elderly, it does occur in young patients as well, and it would be inappropriate to assume that patients with early onset of PD should necessarily be treated the same as patients with older onset of PD. For these reasons and for the possibility of variable and even contradictory results, it would be necessary to classify patients in various groups. We recommend NAC supplements based on scientific rationale for three population sub-groups: (a) persons at high risk for PD, (b) patients with early stage PD and (c) patients on established L-dopa therapy in combination with NAC. These recommendations should be assessed for efficacy in a well-designed clinical trial.

In regard to the oral NAC administration, a minimal dose of 600 mg/day and a maximum of 1800 mg/day would be into the

range of use. It would be important to record whatever adverse effect or intolerance during the treatment and also what is the cause of the treatment interruption.

Conclusions

The present paper shows that many experimental data in the last 10 years support the concept that NAC may be a singular substance for the treatment and prevention of PD and that a well-designed clinical trial is justified.

Conflict of interest statement

No conflict of interest is declared.

References

- [1] Agid Y, Ruberg M, Javoy-Agid F, et al. Are dopaminergic neurons selectively vulnerable to Parkinson's disease? *Adv Neurol* 1993;60:148–64.
- [2] Veldman BA, Wijn AM, Knoers N, et al. Genetic and environmental risk factors in Parkinson's Disease. *Clin Neurol Neurosurg* 1998;100:15–26.
- [3] Williams AC, Smith ML, Waring RH, et al. Idiopathic Parkinson's disease: a genetic and environmental model. *Adv Neurol* 1999;80:215–8.
- [4] Bowling AC, Mutisya EM, Walker LC, et al. Age-dependent impairment of mitochondrial function in primate brain. *J Neurochem* 1993;60:1964–7.
- [5] Curti D, Giangare MD, Redolfi ME, et al. Age-related modification of cytochrome c oxidase activity in discrete brain regions. *Ageing Dev* 1990;55:171–80.
- [6] Ferrández ML, Martínez-Banaclocha M, De Juan E, et al. Impairment of mitochondrial oxidative phosphorylation in the brain of aged mice. *Brain Res* 1994;644:335–8.
- [7] Martínez-Banaclocha M, Ferrández ML, De Juan E, et al. Age-related changes in glutathione and lipid peroxide content in mouse synaptic mitochondria: relationship to cytochrome c oxidase decline. *Neurosci Lett* 1994;170:121–4.
- [8] Martínez-Banaclocha M, Ferrández ML, Diez A, et al. Depletion of cytosolic GSH decreases the ATP levels and viability of synaptosomes from aged mice but not from young mice. *Mech Ageing Dev* 1995;84:77–81.
- [9] Martínez-Banaclocha M, Hernández AI, Martínez N. *N*-acetylcysteine delays age-associated memory impairment in mice. role in synaptic mitochondria. *Brain Res* 2000;855:100–6.
- [10] Martínez-Banaclocha M, Hernández AI, Martínez N, et al. Age-related increase in oxidized proteins in mouse synaptic mitochondria. *Brain Res* 1996;731:246–8.
- [11] Langston JW, Ballard P, Tetrud JW, et al. Chronic parkinsonism in humans due to a product of meperidine-analog synthesis. *Science* 1983;219:979–80.
- [12] Sayre LM. Biochemical mechanism of action of the dopaminergic neurotoxin MPTP. *Toxicol Lett* 1989;48:121–49.
- [13] Langston JW, Forno LS, Tetrud J, et al. Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure. *Ann Neurol* 1999;46:598–605.
- [14] Mizuno Y, Ikebe S, Hattori N, et al. Role of mitochondria in the etiology and pathogenesis of Parkinson's disease. *Biochem Biophys Acta* 1995;1271:265–74.
- [15] Swerdlow RH, Parks JK, Miller SW, et al. Origin and functional consequences of the complex I defect in Parkinson's disease. *Ann Neurol* 1996;40:663–71.
- [16] Schapira AH, Mann VM, Cooper JM, et al. Anatomic and disease specificity of NADH CoQ1 reductase (complex I) deficiency in Parkinson's disease. *J Neurochem* 1990;55:2142–5.
- [17] Gilks WP, Abou-Sleiman PM, Gandhi S, et al. A common LRRK2 mutation in idiopathic Parkinson's disease. *Lancet* 2005;365:415–6.
- [18] Perry TL, Godin DV, Hansen S. Parkinson's disease: a disorder due to nigral glutathione deficiency? *Neurosci Lett* 1982;33:305–10.
- [19] Perry TL, Yong VW. Idiopathic Parkinson's disease, progressive supranuclear palsy and glutathione metabolism in the substantia nigra of patients. *Neurosci Lett* 1986;67:269–74.
- [20] Meister A. Glutathione metabolism and its selective modification. *J Biol Chem* 1988;263:17205–8.
- [21] Jenner P. Oxidative mechanisms in nigral cell death in Parkinson's disease. *Mov Disord* 1998;13:24–34.
- [22] Cassarino DS, Fall CP, Swerdlow RH, et al. Elevated reactive oxygen species and antioxidant enzyme activities in animal and cellular models of Parkinson's disease. *Biochim Biophys Acta* 1997;1362:77–86.
- [23] Jenner P. Oxidative stress in Parkinson's disease. *Ann Neurol* 2003;53:S26–36 [discussion S36–38].
- [24] Clementi E, Brown GC, Feelisch M, Moncada S. Persistent inhibition of cell respiration by nitric oxide: crucial role of S-nitrosylation of mitochondrial complex I and protective action of glutathione. *Proc Natl Acad Sci USA* 1998;95:7631–6.
- [25] Hsu M, Srinivas B, Kumar J, et al. Glutathione depletion resulting in selective mitochondrial complex I inhibition in dopaminergic cells is via an NO-mediated pathway not involving peroxynitrite: implications for Parkinson's disease. *J Neurochem* 2005;92:1091–103.
- [26] Jha N, Jurma O, Lalli G, et al. Glutathione depletion in PC12 results in selective inhibition of mitochondrial complex I activity. Implications for Parkinson's disease. *J Biol Chem* 2000;275:26096–7001.
- [27] Conway KA, Rochet JC, Bieganski RM, et al. Kinetic stabilization of the alpha-synuclein proto-fibril by a dopamine-alpha-synuclein adducts. *Science* 2001;294:1346–9.
- [28] Giasson BI, Duda JE, Murray IV, et al. Oxidative damage linked to neurodegeneration by selective alpha-synuclein nitration in synucleinopathy lesions. *Science* 2000;290:985–9.
- [29] Shults CW, Beal MF, Fontaine D, et al. Absorption, tolerability, and effects on mitochondrial activity of oral coenzyme Q10 in parkinsonian patients. *Neurology* 1998;50:793–5.
- [30] Parkinson Study Group. Effects of tocopherol and deprenyl on the progression of disability in early Parkinson's disease. *New Engl J Med* 1993;328:176–83.
- [31] Etmann M, Gill SS, Samii A. Intake of vitamin E, vitamin C, and carotenoids and the risk of Parkinson's disease: a meta-analysis. *Lancet Neurol* 2005;4:362–5.
- [32] Sechi G, Deledda MG, Bua G, et al. Reduced intravenous glutathione in the treatment of early Parkinson's disease. *Progr Neuropsychopharmacol Biol Psychiatry* 1996;20:1159–70.
- [33] Wang XF, Cynader MS. Pyruvate released by astrocytes protects neurons from copper-catalyzed cysteine neurotoxicity. *J Neurosci* 2001;21:3322–31.
- [34] Martínez-Banaclocha M. *N*-acetylcysteine elicited increase in complex I activity in synaptic mitochondria from aged mice. Implications for treatment of Parkinson's disease. *Brain Res* 2000;859:173–5.
- [35] Martínez-Banaclocha M. Therapeutic potential of *N*-acetyl-cysteine in age-related mitochondrial neurodegenerative diseases. *Med Hypotheses* 2001;56:472–7.
- [36] Martínez-Banaclocha M, Martínez N. *N*-acetylcysteine elicited increase in cytochrome c oxidase activity in mice synaptic mitochondria. *Brain Res* 1999;842:249–51.
- [37] Martínez-Banaclocha M, Hernández AI, Martínez N, et al. *N*-acetylcysteine protects against age-related increase in oxidized proteins in mouse synaptic mitochondria. *Brain Res* 1997;762:256–8.
- [38] Martínez-Banaclocha M, Martínez N, Hernández, et al. Hypothesis: can *N*-acetylcysteine be beneficial in Parkinson's disease? *Life Sci* 1999;64:1253–7.
- [39] Medina S, Martínez-Banaclocha M, Hernanz A. Antioxidants inhibit the human cortical neuron apoptosis induced by hydrogen peroxide, tumor necrosis factor alpha, dopamine and beta-amyloid peptide 1–42. *Free Radic Res* 2002;36:1179–84.
- [40] Pocernich CB, La Fontaine M, Butterfield DA. In-vivo glutathione elevation protects against hydroxyl free radical-induced protein oxidation in rat brain. *Neurochem Int* 2000;36:185–91.
- [41] Vina J, Romero FJ, Saez GT, et al. Effects of cysteine and *N*-acetyl cysteine on GSH content of brain of adult rats. *Experientia* 1983;39:164–5.
- [42] Coccoa T, Sgobbo P, Clemente M, et al. Tissue-specific changes of mitochondrial functions in aged rats: effect of a long-term dietary treatment with *N*-acetylcysteine. *Free Radic Biol Med* 2005;38:796–805.
- [43] Perry TL, Yong VW, Clavier RM, et al. Partial protection from the dopaminergic neurotoxin *N*-methyl-4-phenyl-1,2,3,6-tetrahydropyridine by four different antioxidants in the mouse. *Neurosci Lett* 1985;60:109–14.
- [44] Sharma A, Kaur P, Kumar V, et al. Attenuation of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine induced nigro-striatal toxicity in mice by *N*-acetyl cysteine. *Cell Mol Biol* 2007;53:48–55.
- [45] Offen D, Ziv I, Sternin H, et al. Prevention of dopamine-induced cell death by thiol antioxidants: possible implications for treatment of Parkinson's disease. *Exp Neurol* 1996;141:32–9.
- [46] Nicoletti VG, Marino VM, Cuppari C, et al. Effect of antioxidant diets on mitochondrial gene expression in rat brain during aging. *Neurochem Res* 2005;30:737–52.
- [47] McCann UD, Wong DF, Yokoi F, et al. Reduced striatal dopamine transporter density in abstinent methamphetamine and methcathinone users: evidence from positron emission tomography studies with [¹¹C]WIN-35,428. *J Neurosci* 1998;18:8417–22.
- [48] Wilson JM, Kalasinsky KS, Levey AI, et al. Striatal dopamine nerve terminal markers in human, chronic methamphetamine users. *Nat Med* 1996;2:699–703.
- [49] Hashimoto H, Tsukada H, Nishiyama S, et al. Protective effects of *N*-acetyl-L-cysteine on the reduction of dopamine transporters in the striatum of monkeys treated with methamphetamine. *Neuropsychopharmacology* 2004;29:2018–23.
- [50] Xu J, Kao SY, Lee FJ, et al. Dopamine-dependent neurotoxicity of alpha-synuclein: a mechanism for selective neurodegeneration in Parkinson disease. *Nat Med* 2002;8:600–6.
- [51] Clark J, Clore EL, Zheng K, et al. Oral *N*-acetyl-cysteine attenuates loss of dopaminergic terminals in alpha-synuclein over-expressing mice. *PLoS One* 2010;5:e12333.
- [52] Ferrari G, Yan CYI, Greene LA. *N*-acetylcysteine (D- and L-stereoisomers) prevents apoptotic death of neuronal cells. *J Neurosci* 1995;15:2857–66.
- [53] Mayer M, Noble M. *N*-acetyl-L-cysteine is a pluripotent protector against cell death and enhancer of trophic factor-mediated cell survival in vitro. *Proc Natl Acad Sci USA* 1994;91:7496–500.
- [54] Green DR, Reed JC. Mitochondria and apoptosis. *Science* 1998;281:1309–12.
- [55] Ratan RR, Murphy TH, Baraban JM. Macromolecular synthesis inhibitors prevent oxidative stress-induced apoptosis in embryonic cortical neurons by

- shunting cysteine from protein synthesis to glutathione. *J Neurosci* 1994;17:4385–92.
- [56] Lafon C, Mathiew C, Guerrin M, et al. Transforming growth factor beta 1-induced apoptosis in human ovarian carcinoma cells: protection by the antioxidant *N*-acetylcysteine and bcl-2. *Cell Growth Differ* 1996;7:1095–104.
- [57] Aoki E, Yano R, Yokoyama H, Kato H, Araki T. Role of nuclear transcription factor kappa B (NF-kappaB) for MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine)-induced apoptosis in nigral neurons of mice. *Exp Mol Pathol* 2009;86:57–64.
- [58] Sha D, Chin LS, Li L. Phosphorylation of parkin by Parkinson disease linked kinase PINK1 activates parkin E3 ligase function and NF-kappaB signaling. *Hum Mol Genet* 2010;19:352–63.
- [59] Cossarizza A, Franceschi C, Monti D, et al. Protective effects of *N*-acetylcysteine in tumor necrosis factor-alpha-induced apoptosis in U937 cells: the role of mitochondria. *Exp Cell Res* 1996;220:232–40.
- [60] Talley AK, Dewhurst S, Perry SW, et al. Tumor necrosis factor-alpha-induced apoptosis in human neuronal cells: protection by the antioxidant *N*-acetylcysteine and the genes bcl-2 and crmA. *Mol Cell Biol* 1995;15:2359–66.
- [61] Bagh MB, Maiti AK, Jana S, et al. Quinone and oxyradical scavenging properties of *N*-acetylcysteine prevent dopamine mediated inhibition of Na⁺, K⁺-ATPase and mitochondrial electron transport chain activity in rat brain: implications in the neuroprotective therapy of Parkinson's disease. *Free Radic Res* 2008;42:574–81.